QUALITY CONTROL PROCEDURES (Optional)

I INTRODUCTION

Trypticase™ Soy Agar with 5% Sheep Blood is used for the growth of fastidious organisms and for the visualization of hemolytic reactions. MacConkey II Agar is a selective and differential medium for the detection of coliform organisms and enteric pathogens.

II PERFORMANCE TEST PROCEDURE

A. Trypticase Soy Agar with 5% Sheep Blood

1. Inoculate representative samples with dilutions of the cultures listed below.
   a. Using a volumetric pipettor or equivalent method, deliver 0.1 mL of a dilution yielding 30–300 CFU to each plate and spread-inoculate using a sterile glass spreader.
   b. Incubate the Staphylococcus and Escherichia strains at 35 ± 2 °C in an aerobic atmosphere and the Streptococcus strains at 35 ± 2 °C in an aerobic atmosphere supplemented with carbon dioxide.
2. Examine plates after 18–24 h for growth, colony size and hemolytic reactions.
3. Expected Results

<table>
<thead>
<tr>
<th>CLSI Organisms</th>
<th>ATCC®</th>
<th>Recovery</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Streptococcus pyogenes</em></td>
<td>19615</td>
<td>Growth, beta hemolysis</td>
</tr>
<tr>
<td><em>Streptococcus pneumoniae</em></td>
<td>6305</td>
<td>Growth, alpha hemolysis</td>
</tr>
<tr>
<td><em>Staphylococcus aureus</em></td>
<td>25923</td>
<td>Growth</td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>25922</td>
<td>Growth</td>
</tr>
</tbody>
</table>

*Recommend organism strain for User Quality Control.

B. BBL MacConkey II Agar

1. Inoculate representative samples with dilutions of the cultures listed below.
   a. Streak the plates for isolation using 18- to 24-h broth cultures diluted 10⁻¹. For Proteus mirabilis, make two additional ten-fold dilutions prior to streaking.
   b. Incubate the plates at 35 ± 2 °C in an aerobic atmosphere.
   c. Include Trypticase™ Soy Agar with 5% Sheep Blood plates as nonselective controls for all organisms.
2. Examine plates after 18–24 h for growth, colony size, pigmentation and selectivity.
3. Expected Results

<table>
<thead>
<tr>
<th>CLSI Organisms</th>
<th>ATCC®</th>
<th>Recovery</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Escherichia coli</em></td>
<td>25922</td>
<td>Growth</td>
</tr>
<tr>
<td><em>Proteus mirabilis</em></td>
<td>12453</td>
<td>Growth, inhibition of swarming (partial)</td>
</tr>
<tr>
<td><em>Salmonella enterica</em></td>
<td>14028</td>
<td>Growth</td>
</tr>
<tr>
<td>subsp. enterica</td>
<td></td>
<td>Colorless</td>
</tr>
<tr>
<td>serotype Typhimurium</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Enterococcus faecalis</em></td>
<td>29212</td>
<td>Inhibition (partial)</td>
</tr>
</tbody>
</table>

Additional Organisms

   | Pseudomonas aeruginosa | 10145 | Growth                          |
   | Shigella dysenteriae  | 9361  | Growth                          |

*Recommend organism strain for User Quality Control.

III ADDITIONAL QUALITY CONTROL

1. Examine plates as described under “Product Deterioration.”
2. Visually examine representative plates to assure that any existing physical defects will not interfere with use.
3. Determine the pH potentiometrically at room temperature for adherence to the specification of 7.3 ± 0.2 (TSA II) and 7.1 ± 0.2 (MacConkey II Agar).
4. Note the firmness of plates during the inoculation procedure.
5. Incubate uninoculated representative plates at 35 ± 2 °C for 72 h and examine for microbial contamination.

PRODUCT INFORMATION

IV INTENDED USE

Trypticase™ Soy Agar with 5% Sheep Blood is used for cultivating fastidious microorganisms and for the visualization of hemolytic reactions produced by many bacterial species. MacConkey II Agar is a selective and differential medium for the detection of coliform organisms and enteric pathogens.

V SUMMARY AND EXPLANATION

A. Trypticase Soy Agar with 5% Sheep Blood

The nutritional composition of Trypticase™ Soy Agar has made it a popular medium, both unsupplemented and as a base for media containing blood. Trypticase™ Soy Agar with 5% Sheep Blood is extensively used for the recovery and cultivation of fastidious microbial species and for the determination of hemolytic reactions which are important differentiating characteristics for bacteria, especially Streptococcus species.
B. MacConkey II Agar

At the present time, many culture media are available to the laboratorian for the isolation, cultivation and identification of enteric bacteria. One of the earliest of these was developed by MacConkey and first described as a brief published note. The landmark paper on MacConkey Agar was published in 1905 and contained detailed descriptions of the medium and the bacterial growth patterns obtained. This formulation was devised in the knowledge that bile salts are precipitated by acids and certain enteric microorganisms ferment lactose whereas others do not possess this ability. Since the publication of the early papers, the MacConkey Agar formula has been modified many times. A compilation of culture media published in 1930 lists ten modifications which were published up to that time. More recent modifications include use of additives (e.g., kanamycin) and the deletion of certain ingredients (e.g., crystal violet, and neutral red). MacConkey Agar is recommended for use with clinical specimens likely to contain mixed microbial flora, such as urine, respiratory and wound, because it allows a preliminary grouping of enteric and other gram-negative bacteria. It is also utilized in the microbiological examination of foods.

The BBL MacConkey II Agar formulation was made available in 1983. It was specially designed to improve the inhibition of swarming Proteus species, to achieve more definitive differentiation of lactose fermenters and nonfermenters, and for the promotion of superior growth of enteric pathogens.

VI PRINCIPLES OF THE PROCEDURE

A. Trypticase Soy Agar with 5% Sheep Blood

The combination of casein and soy peptones in the Trypticase Soy Agar base render the medium highly nutritious by supplying organic nitrogen, particularly amino acids and larger-chained peptides. The sodium chloride maintains osmotic equilibrium. Defibrinated sheep blood is the most widely used blood for enriching agar base media. Hemolytic reactions of streptococci are proper and growth of Haemophilus hemolyticus, a nonpathogen whose hemolytic colonies are indistinguishable from those of beta-hemolytic streptococci, is inhibited.

Trypticase Soy Agar with 5% Sheep Blood (TSA II) provides excellent growth and beta hemolysis by Streptococcus pyogenes (Lancefield group A) and also provides excellent growth and appropriate hemolytic reactions with other fastidious organisms. It is suitable for use with low concentration (0.04 unit) bacitracin discs (Taxo™ A) for presumptive identification of group A streptococci (S. pyogenes).

B. MacConkey II Agar

MacConkey II Agar is a selective and differential medium. It is only slightly selective since the concentration of bile salts, which inhibits gram-positive microorganisms, is low in comparison with other enteric plating media. Crystal violet also is included in the medium to inhibit the growth of gram-positive bacteria, especially enterococci and staphylococci.

Differential of enteric microorganisms is achieved by the combination of lactose and the neutral red indicator. Colorless or pink to red colonies are produced depending upon the ability of the isolate to ferment the carbohydrate.

VII REAGENTS

**Trypticase Soy Agar with 5% Sheep Blood (TSA II)**

Approximate Formula* Per Liter Purified Water

<table>
<thead>
<tr>
<th>Component</th>
<th>Weight</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pancreatic Digest of Casein</td>
<td>14.5 g</td>
</tr>
<tr>
<td>Trypticase</td>
<td>14.0 g</td>
</tr>
<tr>
<td>Sodium Chloride</td>
<td>5.0 g</td>
</tr>
<tr>
<td>Bile Salts</td>
<td>0.15 g</td>
</tr>
</tbody>
</table>

*Adjusted and/or supplemented as required to meet performance criteria.

**MacConkey II Agar**

Approximate Formula* Per Liter Purified Water

<table>
<thead>
<tr>
<th>Component</th>
<th>Weight</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pancreatic Digest of Gelatin</td>
<td>17.0 g</td>
</tr>
<tr>
<td>Sodium Chloride</td>
<td>5.0 g</td>
</tr>
<tr>
<td>Peptic Digest of Animal Tissue</td>
<td>0.15 g</td>
</tr>
<tr>
<td>Lactose</td>
<td>10.0 g</td>
</tr>
<tr>
<td>Bile Salts</td>
<td>0.15 g</td>
</tr>
</tbody>
</table>

*Adjusted and/or supplemented as required to meet performance criteria.

**Warnings and Precautions: For in vitro Diagnostic Use.**

If excessive moisture is observed, invert bottom over an off-set lid and allow to air dry in order to prevent formation of a seal between the top and bottom of the plate during incubation.

Pathogenic microorganisms, including hepatitis viruses and Human Immunodeficiency Virus, may be present in clinical specimens. “Standard Precautions” and institutional guidelines should be followed in handling all items contaminated with blood and other body fluids. After use, prepared plates, specimen containers and other contaminated materials must be sterilized by autoclaving before discarding.

**Storage Instructions:** On receipt, store plates in the dark at 2–8 °C. Avoid freezing and overheating. Do not open until ready to use. Minimize exposure to light. Prepared plates stored in their original sleeve wrapping at 2–8 °C until just prior to use may be inoculated up to the expiration date and incubated for recommended incubation time. Allow the medium to warm to room temperature before inoculation.

**Product Deterioration:** Do not use plates if they show evidence of microbial contamination, discoloration, drying or other signs of deterioration.

**VIII SPECIMEN COLLECTION AND HANDLING**

A variety of swabs and containers have been devised for collecting specimens. Specimens should be obtained before antimicrobial therapy has been administered. Provision must be made for prompt delivery to the laboratory. Several holding media or transport systems, such as BBL specimen collection and transport products, have been devised to prolong the survival of microorganisms when a significant delay is expected between collection and definitive culturing.

Refer to appropriate texts for details of specimen collection and handling procedures.

The laboratory must be furnished with sufficient clinical information to enable the microbiologist to select the most suitable media and appropriate techniques.
IX PROCEDURE

Material Provided: Trypticase Soy Agar with 5% Sheep Blood (TSA II) and MacConkey II Agar (I Plate)

Materials Required But Not Provided: Ancillary culture media, reagents, quality control organisms and laboratory equipment as required.

Test Procedure: Observe aseptic techniques.

The agar surface should be smooth and moist, but without excessive moisture. Streak the specimen as soon as possible after it is received in the laboratory. The streak plate is used primarily to isolate pure cultures from specimens containing mixed flora. Alternatively, if material is being cultured directly from a swab, roll the swab over a small area of the surface at the edge; then streak from this inoculated area.

Incubate plates, protected from light, at 35 ± 2 °C for 18–24 h. With respiratory specimens, incubate in an aerobic atmosphere supplemented with carbon dioxide. With other specimens, incubate aerobically without added CO₂.

User Quality Control: See “Quality Control Procedures.” Each lot of media has been tested using appropriate control organisms and this testing meets product specifications and CLSI standards, where relevant. As always, QC testing should be performed in accordance with applicable local, state, federal or country regulations, accreditation requirements, and/or your laboratory’s standard quality control procedures.

X RESULTS

After incubation most plates will show an area of confluent growth. Because the streaking procedure is, in effect, a “dilution” technique, diminishing numbers of microorganisms are deposited on the streaked areas. Consequently, one or more of these areas should exhibit isolated colonies of the organisms contained in the specimen. Further, growth of each organism may be semi-quantitatively scored on the basis of growth in each of the streaked areas.

Typical results on Trypticase Soy Agar with 5% Sheep Blood are as follows:

1. Hemolytic streptococci may appear as translucent or opaque, grayish, small (1 mm), or large matt and mucoid (2–4 mm) colonies, encircled by a zone of hemolysis. Gram stains should be made and examined to check the macroscopic findings. (Other organisms which may cause hemolysis include Listeria, various corynebacteria, hemolytic staphylococci, Escherichia coli and Pseudomonas.) In reporting, approximate quantitation of the number of colonies of hemolytic streptococci may be helpful to the clinician.

2. Pneumococci usually appear as very flat, smooth, translucent, grayish and sometimes mucoid colonies surrounded by a narrow zone of “green” (alpha) hemolysis.

3. Staphylococci appear as opaque, white to gold-yellow colonies with or without zones of beta hemolysis.

4. Listeria. Small zones of beta hemolysis are produced. They may be distinguished by their rod shape in stains, and by motility at room temperature.

5. Other organisms representing minimal flora and clinically significant isolates can also be expected to grow on this nonselective formulation.

Typical colonial morphology on MacConkey II Agar is as follows:

E. coli .......................................................... Pink to rose-red (may be surrounded by a zone of precipitated bile)

Enterobacter/Klebsiella ......................... Mucoid, pink

Proteus ...................................................... Colorless, swarming in areas of isolated colonies is inhibited

Salmonella ................................................. Colorless

Shigella ..................................................... Colorless

Pseudomonas ........................................... Irregular, colorless to pink

Gram-positive bacteria ......................... No growth to slight growth

XI LIMITATIONS OF THE PROCEDURE

It has been reported that some Enterobacteriaceae and Pseudomonas aeruginosa are inhibited on MacConkey Agar when incubated in a CO₂-enriched atmosphere.¹⁵

Not all strains of E. coli ferment lactose.

Some diagnostic tests may be performed with the primary plate. However, a pure culture is recommended for biochemical tests and other identification procedures. Consult appropriate texts for detailed information and recommended procedures.⁵ ¹⁵-¹⁹

A single medium is rarely adequate for detecting all organisms of potential significance in a specimen. It should be recognized that organisms generally susceptible to the antimicrobial agent in a selective medium may be completely or only partially inhibited depending upon the concentration of the agent, the characteristics of the microbial strain and the number of organisms in the inoculum. Organisms that are generally resistant to the antimicrobial agent should not be inhibited. Cultures of specimens grown on selective media should, therefore, be compared with specimens cultured on nonselective media to obtain additional information and help ensure recovery of potential pathogens.

XII PERFORMANCE CHARACTERISTICS

Trypticase Soy Agar with 5% Sheep Blood

Trypticase Soy Agar with 5% Sheep Blood was used as a control in a study using broth-enhanced culture (Todd Hewitt) and Optical Immunoassay method for the diagnosis of β-hemolytic streptococcal infection. Five hundred two (502) specimens were tested. TSA with 5% Sheep Blood had a sensitivity and specificity of 92.5% and 99.4%, respectively.²⁰

Nguyen et al. used Trypticase Soy Agar with 5% Sheep Blood as the “gold standard” for the detection of group B Streptococcus from the lower genital tract of pregnant women.²¹ In another study, Rossmann et al. successfully reisolated Lautropia mirabilis on Trypticase Soy Agar with 5% Sheep Blood from the oral cavities of human immunodeficiency virus infected children.²² Of the 85 children evaluated in this study, 35 (41.4%) were positive for L. mirabilis. Isenberg et al. used Trypticase Soy Agar with 5% Sheep Blood as a control to evaluate the recovery of Enterococcus from a selective medium under study.²³ Two hundred fifty (250) group D streptococcal strains isolated from clinical material and 8 strains obtained from the National Communicable Disease Center (Atlanta, Ga.) were used.
### MacConkey II Agar

Prior to release, all lots of MacConkey II Agar are tested for performance characteristics. Representative samples of the lot are streak-inoculated with the following cultures: *Escherichia coli* (ATCC 25922), *Proteus mirabilis* (ATCC 12453), *Pseudomonas aeruginosa* (ATCC 10145), *Salmonella Typhimurium* (ATCC 14028), *Shigella dysenteriae* (ATCC 9361) and *Enterococcus faecalis* (ATCC 29212). The inoculum for *E. faecalis* is diluted to yield $10^4 – 10^5$ colony-forming units (CFU) per plate; the inocula for all other organisms is diluted to yield $10^2 – 10^3$ CFU/plate. After inoculation, the plates are incubated at $35 ± 2 ^\circ C$ in an aerobic atmosphere. After 18–24 h incubation, colonies of *E. coli* are rose-red and may be surrounded by precipitated bile; *P. mirabilis* exhibits fair to heavy growth of colorless colonies and swarming of the colonies is inhibited; *P. aeruginosa* shows areas of confluent growth which may exhibit green to yellow-green pigmentation while individual colonies show pink to green pigmentation; *Salmonella Typhimurium* gives fair to heavy growth of colorless colonies; *S. dysenteriae* shows growth of colorless to pink colonies; *E. faecalis* is completely to partially inhibited (fair growth) and the colonies may be pink in color.

### XIIV AVAILABILITY

<table>
<thead>
<tr>
<th>Cat. No.</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>221290</td>
<td>BD BBL™ Trypticase™ Soy Agar with 5% Sheep Blood (TSA II), and MacConkey II Agar I Plate™, Pkg. of 20 plates</td>
</tr>
<tr>
<td>221291</td>
<td>BD BBL™ Trypticase™ Soy Agar with 5% Sheep Blood (TSA II), and MacConkey II Agar I Plate™, Ctn. of 100 plates</td>
</tr>
</tbody>
</table>

### XIV REFERENCES


Technical Information: In the United States contact BD Technical Service and Support at 800-638-8663 or www.bd.com/ds.